# **Rearrangement Processes in the Bicycle [3.2.1] heptane Moiety of 12,17-Bifunctionalized ent=Beyer-Senes**

Andrés Garcia-Granados<sup>\*</sup> and Andrés Parra

**Departamento de Química Orgánica Pa&ad de Ciencias. Universidad de Granada 18071. Granada.** Spain.

*(Received in UK 18 June* 1991)

**Z&y Worrls: Diterpenoids, ent-beyer-lSenes, ent-atis-l3-ems,** *rearrangements,* **substituyent effects.** 

Abstract: Several rearrangements of 17-mesyloxy and 12,17-dimesyloxy derivatives of ent-beyer-15-enes were carried out. *Reamzngement processes mainly occurred with :he pmiiciipanion* of the functional *gmup ot C-12, although the gmup at C-17ond the C-IS/C-l6 double bond aided in on observed 16(13)-12/11(12) -13 double twmangement. The influence of the gmups at C-12 and their stereochemisby, of the meghy pups at C-17 and of the C-IS/C-l6 double bond is studied and the stereochemistry of the observed rearrangement processes is discussed.* 

## **INTRODUCTION**

The rearrangement of tetracyclic diterpenoids has been extensively studied in the last two decades. In general, the rearrangements studied in solvolytic reactions<sup>15</sup> or from epoxy compounds<sup>612</sup> were carried out on starting material with only one functional group present in the moiety of the molecule capable of undergoing rearrangement. On some occasions, the double bond originally present in the natural skeleton was hydrogenated before setting up the rearrangement experiment. However, we think that the presence of other functional groups near the leaving group may have a decisive quali-or/and quantitative influence on the course of the reaction, as reported in the conversion of  $ent$ -beyer-15-ene to  $ent$ -atis-13ene<sup>13,14</sup>, in the ruthenium-catalyzed rearrangement of epoxybeyeranes functionalized at  $C^{-14^{15}}$  or in the acid-catalyzed rearrangement of the  $ent$ -beyer-15-en-12-one system<sup>16</sup>. In this work we present the results of the rearrangement of ent-beyer-15enes functionalized at C-12 and C-17 to study the influence of the presence/absence of functional groups in these positions, principally of the C-15/C-16 double bound, and to compare the results with those described by McAlees and McCrindle in their general scheme of rearrangement of tetracyclic diterpenoids functionalized at C-12'.

# **RESULTS AND DISCUSSION**

Treatment of ent-1ß-acetoxy-12 $\alpha$ ,17-dihydroxybeyer-15-ene (1-acetyljativatriol,1<sup>17</sup>) with mesyl chloride (M&l) at room temperature yielded its 17-mesyl derivative (2,90%) which afterward was treated with AcOK/AcOH for 24 hours under reflux (see Experimental part), after which starting material (2, 15%), 1,17-diacetyljativatriol<sup>18</sup> (3, 25%) and a rearranged product (4, 40%) were isolated (Scheme I). This rearranged product (4), the main product of this process, was a triacetyl derivative (MS, see Experimental part; <sup>1</sup>H NMR, Table I; <sup>13</sup>C NMR, Table III), with one acetoxymethylene (situated at a quaternary carbon) and two acetoxymethine groups. One of these can be attributed to the original group at C-l of the unaltered A ring, but the other one cannot be situated at C-12 of an ent-beyer-15-ene system. In the new skeleton, this acetoxymethine group, with a <sup>1</sup>H NMR signal at  $\delta$  4.98 (1H, d, J=3.9 Hz), was allylic to a double bond, which was proved by double resonance experiments. Moreover, the <sup>13</sup>C NMR chemical shift of C-9 was very different ( $\delta$  40.50) to that of ent-beyer-15-ene products ( $\delta$  49.32 for product 2).

## SCHEME I



Solvolysis of ent-12 $\alpha$ -tosyloxybeyerane, described by Coates & Bertram<sup>3</sup> and McAlees & McCrindle<sup>4</sup>, gave ent-atisane. ent-Atisene compounds were also obtained by rearrangements of 12,17isopropylidene or 12,17-thiocarbonyl derivatives of 1-acetyljativatriol<sup>14</sup>. However, the skeleton now obtained was not an ent-atisene, because four non-oxygenated quaternary carbons and two non-oxygenated methine carbons were present in the <sup>13</sup>C NMR spectra of 4, and irradiation of the proton geminal to the allylic acetoxy group transformed the signal of the olefin H-15 ( $\delta$  5.54, dd, J<sub>1</sub>=9.5, J<sub>2</sub>=3.9Hz) into a doublet  $(J=9.5 \text{ Hz})$ .

The configuration at C-12 can be easely. deduced by consideration of the coupling constant of its proton  $(J=3.9 \text{ Hz})$  and the dihedral angles for the two possible configurations calculated after the observation of Dreiding models (40 and 75" respectively). Taking into account the influence of the electronegativity of the acetoxy group on the coupling constant, its observed value (3.9 Hz) was in accord with an R configuration (near 4 Hz is described<sup>19</sup>), whereas for the S configuration a coupling constant near zero was expected. In addition, C/H correlation experiments were performed to assign the chemical shifts of protons at C-11 and C-15. Subsequent n.0.e difference experiments desmonstrated the spatial proximity between H-12 and ent-11ß-H, H-16 and the C-20 methyl group and between ent-11 $\alpha$ -H and the acetoxymethylene group of C-17. Thus, product 4 must be the result of a  $16(13) \rightarrow 12/11(12) \rightarrow 13$  double rearrangement process. This structure was proposed as a possibility in the general rearrangement scheme of 12 $\alpha$  and 12B-hydroxybeyeranes by McAlees et all<sup>5</sup>, but not isolated. Thus, the hydroxyl group at C-12 is likely to participate in this process, as shown in Chart I.

### CHART I



To prove the participation of the hydroxyl group at C-12, we acetylated the starting material 2 to obtain the diacetyl derivative 5, which was treated under the same conditions as 2. In this case, the rearranged product 4 was also obtained, but the yield was only 20%. The product of acetolysis at C-17 was the main one isolated from this process. Moreover, starting product (5, 10%) and the atisene products  $7^{13}(15%)$  and  $8^{13}(10%)$  were isolated (Scheme I). In this case, solvolysis of the mesyloxy group at C-17 and participation of the acetoxy group at C-12,as indicated in Chart II, seemed reasonable. The main process

in the reaction was the direct acetolysis at C-17 (via a) and the more classical rearrangement toward entatis-13-ene compounds (via c) was more extensive than the double rearrangement (via b). It seems clear that the rearrangement of 2 and 5 could occur (except for product 6), with the participation of both groups at C-12 and C-17.

# CHART II



To prove if the process was initiated at C-12, we mesylated product 1 at reflux to obtain a dimesylate, that was not isolated. In this case, the atisene compounds 10(40%), **11(25%)** and 12(14%) were isolated (Scheme II). The structures of **10** and **11 can** be deduced from comparison with the data of enr-atis-13,16-diene compounds previously described". Products **10** and **11** were unstable and evolved to aldehyde compounds.

## SCHEME II



The structure of product I2 can be deduced from comparison with the data of its 16-epimer compound 13, obtained by mesylation of the natural product 14 (1-acetylsideritol<sup>13</sup>). The <sup>13</sup>C NMR data of both epimer products (12 and 13) were, as expected, very similar with the exception of C-13, C-14 and C-15, and especially C-16 and C-17 (see Table II). As can be seen, all the products (10-12) isolated in this process had an ent-atisene skeleton, and maintained the original mesyloxy group at C-17.

The oxidation of 1 gave ketone  $15<sup>14</sup>$  (Scheme III), which was mesylated to give 16. This product, after 35 hours at reflux under conditions of acetolysis, was recovered unaltered. Hence, for this type of process, the participation of a 12-oxy group seems to be necessary.

### SCHEME III



To study the influence of the stereochemistry of the hydroxyl group at C-12, on the process, ketone 15 was reduced to give ent-128 (17,  $70\%$ <sup>14</sup>) and ent-12 $\alpha$ -hydroxyl derivatives (1, 30%) (Scheme III). Mesylation of 17 at room temperature produced dimesyl derivative 18, which was acetolysed under reflex for 24 hours in a form similar to that described for mesylate 2. In this manner, rearranged products 19  $(40\%)$ , 20  $(20\%)$ , 21  $(15\%)$  and 22  $(15\%)$  were obtained. All these products  $(19-22)$  showed similar <sup>1</sup>H and <sup>13</sup>C NMR spectra (Tables I, III and IV), although with the difference of an acetoxy group (19 and 20) or a mesyloxy group (21 and 22). The skeleton of all these four products was the result of the expected rearrangement toward ent-14(13)-12-beyerene compounds<sup>5</sup>, although with a 13(17),15-diene system, previously described". All four of these products maintained one function at C-17: an acetoxy group in the case of 19 and 20 and a mesyloxy group in the case of products 21 and 22. We carried out a series of n.O.e.-difference experiments on products 19 and 20 to elucidate the configuration at C-13 of the pairs of compounds. Irradiation at H-17 of 19 gave positive n.O.e. with H-12 (6 2.66, m,  $w<sub>w</sub> = 9$  Hz). However, similar irradiation on H-17 of product 20 did not give n.O.e. with H-12, but did give such an effect with H-16 ( $\delta$  6.23, d, J = 11Hz). Thus, products 19 and 21 are E-isomers of 20 and 21 respectively (Z-isomers).

**Evidently, the** configuration at C-12 was decisive in the **development of** the reaction. Thus, the starting material with  $ent$ -12 $\alpha$  functions evolved toward the two-facedly rearranged compounds. On the other hand, the mesylation at reflux of ent-12a-hydroxy compound **(1)** yielded ent-atis-13,16-diene compounds. As the *ent*-12B-hydroxybeyerane and *ent*-12B-hydroxybeyer-15-ene compounds readily evolved toward ent-atisane<sup>3</sup> or ent-atisene compounds<sup>13,14</sup>, the formation of di-abeo compounds such as 4 may well have occurred via the ent-atis-13-ene compound. To prove this hypothesis we performed the acetolysis of 17-mesyl derivative 13 of the *ent*-atis-13-ene natural product  $14^{13}$  (Scheme IV) under the same conditions as described for the acetolysis of 2.In this way, only product 4 (80%) was isolated. Hence, it seems clear that bicyclo[2.2.2]octane partial structure may have provided the means for the formation of products like 4 from ent-beyer-15-ene compounds, but only from starting material without an original leaving group situated at C-12 in ent-beyer-15-ene, which principally evolved towards ent-atis-13,16-diene compounds.

#### SCHEME Iv



Similar behavior was described for the ent-atis-13-ene compound functionalized at C-16 (but not in C-17) which yielded, by treatment with MsCl, ent-13,16-diene compounds independent of its configuration at C-16<sup>20</sup>. However, it is reasonable to think that the rearrangement to 11(12) $\rightarrow$ 13-abeo compound from ent-atis-13-ene compounds may be favored by a 16-era leaving group or by exe-assistance at C-16 by the mesyloxy group present at C-17. To test the influence of the configuration of the possible intermediate in this rearrangement at C-16, we oxidized the natural *ent*-atis-13-ene 14 with NaIO<sub>4</sub> to give ketone  $23^{20}$  which was then treated with NaBH<sub>4</sub> to give ent-16R, 17-nor-atis-13-ene derivative 24 (65%) and its 16S-epimer 25 (35%) (Scheme IV). The configurations at C-16 of both epimers were determined by the 13C NMR chemical shift of C-11 (6 26.93 and 22.66 for 24 and 25 respectively, see Table IV) and, after the assignment pf the chemical shift to the protons at C-11 (C/H correlation, COSY), by one and **two**  dimensional n.O.e. experiments. Thus, we proved that H-16ß of alcohol 24 was spatially near to H-11ß (a 1.38). The H-16 of its epimer compound 25, was near the latter's H-13 (see Table II). However, the main result of the reduction of ketone 23 was the *exo*-epimer 24. Mesylation of 24 at room temperature gave, after 24 hours of reaction, products 26 (95%).

The spectroscopic behavior of 26 indicated that this product was a 17-nor-derivative of 4, although without an acetate group at C-12. Several homo-decoupling experiments indicated that the new hydroxyl group at C-12 ( $\delta$  3.74) was coupled to H-13 ( $J = 3.3$  Hz) and that H-16 was coupled in addition to H-12, with H-15 ( $J=9.6$  Hz) and H-13 (W coupling,  $J=1.9$  Hz). The configuration at C-12 was determined as S on the basis of the value of the H-12/H-13 coupling constant and several n.O.e.-difference experiments which carried out after identification of the chemicals shifts of the protons by C/H correlation. Thus, irradiation at  $\delta$  3.74 (H-12) gave positive n.O.e. for H-13, H-16 and ent-118-H ( $\delta$  0.96, 1H, ddd, J<sub>1</sub>=14,  $J_2 = 9$ ,  $J_3 = 2$  Hz). On the other hand, irradiation at H-14 gave a clear n.O.e with 3H-20, which confirmed stereochemical characteristics of C and D rings of this molecule. Thus, the rearrangement of  $16$ - $\alpha$ o hydroxyl compound 24 gave  $11(12)$  +13-abeo compounds. However, the *endo* isomer 25 gave the *ent*-17-nortrachylobane derivative 27 in the same mesylation reaction as described for 24, although slowly (48 hours) (Scheme IV). The proton geminal to the hydroxyl group at C-14 gave a singlet signal at  $\delta$  4.11 (1H, bs). The <sup>13</sup>C NMR spectrum of 27 gave three new methine groups at  $\delta$  17.23 (two) and 24.85 (one methine group). C/H correlation indicated overlapping of the methine group at  $\delta$  17.23, and correlation with protons at  $\delta$  0.96, 1.32 and 1.38. The configuration at C-14 was determined by n.O.e-difference experiments, as irradiation at H-14 produced positive n.0.e. with one of the previously described protons (6 1.38) and an excellent n.0.e with the methyl group of C-20 (and vice versa), which indicated an 14-R configuration in this compound 27.

To assign the chemical shifts of the cyclopropane moiety and the possible hydride shift in the rearrangements of 24 and 25, respectively, we reduced ketone 23 with NaBD, to give the 16-deuterated compounds 28 and 29 (exo and endo respectively) (Scheme IV). The treatment of 28 with MsCl under the same conditions as described for 24, gave only one compound 30 (Scheme IV), whose spectroscopic studies confirmed the structure proposed for 26. The treatment of 29, as described for 25, gave only product 31 (Scheme IV), whose <sup>13</sup>C NMR spectrum indicated that only the C-16 carbon ( $\delta$  17.23) was deuterated. C/H correlation thus allowed us to assign the chemical shift  $\delta$  1.32 to H-16, and the n.O.e.difference experiment described above, indicated a chemical shift  $\delta$  1.38 to H-13 (and  $\delta$  24.85 to C-13); hence we assigned  $\delta$  0.96 and  $\delta$  17.23 to H-12 and C-12 respectively.

It was therefore evident that the configuration at C-16 and the assistance of the mesyloxy group at C-17 were decisive in determining the skeleton obtained. The 17-mesyloxy group must act with an exeassistance at C-16 of the *ent*-atis-13-ene skeleton in the  $11(12) \rightarrow 13$ -abeo rearrangement. However, when the leaving group was a 16-endo mesyloxy group, ent-atis-13,16-diene (10, 11) and ent-17-nor-trachylobane (product 27) were obtained.

In addition to obtaining di-abeo compounds (like 4) by acetolysis processes, we found that treatment of starting material 2 with diluted HCl at room temperature during a few minutes produced monorearranged (ent-atisene compound 13,25%) and two-facedly rearranged product (32, 15%) (Scheme II). Treatment under such conditions of ent-beyerene **(1)** and enl-atisene (14) compounds did not produce any transformation.

Product 33 was obtained from compound 1 by chlorination at  $C-17$  with  $CCl<sub>a</sub>/PPh<sub>1</sub>$  and by treatment of 17-chloro derivative with tri-n-butyltin hydride<sup>23,24</sup> to give the 17-deoxy compound 33 (Scheme V). This product 33, by treatment with MsCl at room temperature for 24 hours, gave the  $ent$ -atis-13,16diene 34 (main product<sup>20</sup>, 60%), the ent-atis-13,15-diene (35<sup>20</sup>, 20%) and the ent-1B-acetoxy-17-mesyloxy-16S-atis-13-ene 36 (15%). The chemical shift of C-9 of product 36 was characteristic of ent-atis-13-ene compounds, and the observed deshielding of C-11 indicated that the mesyloxymethylene group at C-16 was exo (for both see Table IV). This process was parallel with that described for ent-12 $\alpha$ -tosyl derivatives<sup>3</sup>, with the exception of product 36, in which a functionalization at the methyl group of C-17 was produced. This possibility was not considered in the case of rearrangement of saturated compounds<sup>3</sup>. Presumably, product 36 could be formed through the diene 34 after protonation of the C-13/C-14 double bond, the formation of trachylobane-like structure, mesylation at C-17 and the formation of ent-atis-13-ene compound with the possible assistance of the mesyloxy group at C-17.

#### SCHEME v



In the light of these findings, the presence of a mesyloxy group (or another one as an acetoxy group, as explained in Chart II) at C-17 was decisive, this group assisting at the exe-face at C-16 of the rearranged ent-beyer-15-ene or ent-atis-13-ene. We think that the exo-assistance, or the presence of 16-exogroup (as in 24 or 28), is necessary, although the formation of products like 4 was suggested through 16 endo intermediates on occasion of the study of the cleavage of the cyclopropane ring of ent-trachylobane compounds with thallic acetate<sup>25</sup>. Moreower, to test the influence of the double bond of the starting *ent*beyer-15-ene we hydrogenated the starting material (2) to give product 37, which was acetolyzed under the same conditions as 2. In this form, in addition to starting material 37 (15%), the triacetyl derivative 38 (80%) was isolated (Scheme V).

We therefore think that the presence of double bond was necessary to stabilize the carbocation at C-12 of di-abeo compounds like compound 4, in a parallel form to described in the acid rearrangement of ent-12-ketobever-15-ene system<sup>16</sup>. The new acetoxy group at C-12 of product 38 indicated that the mesyloxy group at C-17 also takes part in the process, but the impossibility of the above mentioned carbocation forming in detectable amounts ruled out evolution toward di-abeo compounds.

The results obtained in this work suggest that the study of rearrangements of unsaturated and/or polyfunctionalized tetracyclic diterpenes would yield further valuable data.

#### **TABLE I**



#### **TAME** II





#### **TABLEIV**



### **EXPERIMENTAL. SECTION**

Melting points (Kofler apparatus) are uncorrected. The NMR spectra (300 MHz 'H and 75.47 MHz  $^{13}$ C) were obtained with a Bruker AM--300 spectrometer equipped with a process controller and an array processor. Samples were dissolved in CDCl<sub>1</sub>. The assignments of <sup>13</sup>C chemical shifts were made with the aid of distortionless enhancement by polarization transfer (DEPT) using a flip angle of 135". Monodimensional n.O.e.-difference experiments were performed by irradiation for 4 s in series of 8 scans with alternate on-resonance and off-resonance. Bruker's programs were used for COSY (45°), NOESY, CONOESY (90 $^{\circ}$ ) and C/H correlation. The optical rotations were measured on a Perkin-Elmer 240 polarimeter. IR spectra were recorded in a FT-IR-Nicolet 20 SX spectrometer. Mass spectra were obtained on a Hewlett-Packard 5988-A spectrometer (CI,methane). Silica gel SDS 60 Å CC (40-60  $\mu$ m) was used for flash chromatography. The colums were pressurized to 0.1 atm with air. CH<sub>2</sub>Cl<sub>2</sub> or CHCl<sub>3</sub> containing increasing amounts of acetone was used as eluent. Analytical plates (silica gel Merck G) were visuallized by spraying with  $H_2SO_4$ -AcOH, followed by heating at 120 $^{\circ}$ C for 5 min.

# GENERAL MESYLATION PROCEDURE.-

Product (30 mg) was dissolved in 2 mL of pyridine and treated with MsCl (0.01 mL) at room temperature for 24 h or under reflux for 4 h. After this time, the reaction products were dropped in a solution of 50 mL of KHSO<sub>4</sub> (10%), extracted with 50 mL of CH<sub>2</sub>Cl<sub>2</sub>, dried over anhydrous NaSO<sub>4</sub> and the solvent was evaporated at reduced pressure.

GENERAL ACETOLYSIS PROCEDURE.-

Mesylate (50 mg) was dissolved in 2 mL AcOK/AcOH (0.5 N), the mixture refluxed for 24 h and then neutralized with NaHCO<sub>3</sub>, extracted with CH<sub>2</sub>Cl<sub>2</sub>, dried over anhidrous Na<sub>2</sub>SO<sub>4</sub> and the solvent evaporated.

MESYLATION OF ent-1B-ACETOXY-12a-DIHYDROXYBEYER-15-ENE (1).-

Product **1** (200 mg) isolated according to reference 22 treated with MsCl as previously described above, gave, after CC, ent-1l3-acetoxy-17mesyloxy-12a-hydroxybeyer-15-ene (2) (180 mg, 90%): Colorless gum;  $[\alpha]_D$  + 18° (CHCl<sub>3</sub>, c 1); IR  $\mu_{\text{max}}$ (CHCl<sub>3</sub>): 3530, 2945, 1718 and 1174 cm<sup>-1</sup>; <sup>1</sup>H NMR: see Table I; <sup>13</sup>C NMR: see Table III; MS, m/z (%): 441 ( $[M+1]^2$ , 2 %), 285 (60), 363 (100). Anal. Calcd for  $C_2Q_6H_4S$ : C, 62.73; H, 8.18. Found: C, 62.80; H, 8.35.

# ACETOLYSIS OF PRODUCT 2.-

Mesylate 2 (150 mg) was treated with AcOK/AcOH as described above under General Acetolysis Procedure to give, after CC: *ent-* 18,17-diacetoxy-12 $\alpha$ -hydroxybeyer-15-ene (3) (37 mg, 25%), *ent-* 18,12 $\alpha$ ,17triacetoxy-16(13)-12,11(12)-13-diabeobeyer-15-ene (4) (60 mg, 40%) and the starting product (2) (23 mg, 15%). Product 3: known product<sup>18</sup>. Product 4: mp : 87-89°C;  $[\alpha]_D$  -12° (CHCl<sub>3</sub>, c 1); IR  $\mu_{max}$  (CHCl<sub>3</sub>): 2948, 1737 and 1246 cm<sup>-1</sup>; <sup>1</sup>H NMR: see Table I; <sup>13</sup>C NMR: see Table III; MS, m/z (%): 447 ( $[M+1]$ <sup>+</sup>, 1%), 387 (100), 327 (20). Anal. Calcd for C<sub>26</sub>O<sub>6</sub>H<sub>38</sub>: C, 69.96; H, 8.52. Found: C, 69.88; H, 8.48. ACETYLATION OF PRODUCT 2.-

Acetylation of 300 mg of product 2 with 7 mL Ac<sub>2</sub>O and 14 mL pyridine at r.t. for 48 h, gave ent-18,12 $\alpha$ -diacetoxy-17-mesyloxybeyer-15-ene (5) (255 mg, 85%) as a colorless gum;  $[\alpha]_D$  +43° (CHCl<sub>3</sub>, c 1); IR  $\mu_{\text{max}}$  (CHCl<sub>3</sub>): 2945, 1720 and 1170 cm<sup>-1</sup>; <sup>1</sup>H NMR: see Table I; <sup>13</sup>C NMR: see Table III; MS, m/z (%); 483 ( $[M+1]^{\dagger}$ , 1%), 363 (40), 423 (100). Anal. Calcd for  $C_2O_2H_{18}S$ : C,62.24; H, 7.88. Found: C, 61.87; H, 8.20.

## ACETOLYSIS OF PRODUCT 5.-

Treatment of 200 mg of product 5 with AcOK/ACOH in the usual manner gave: (4) (40 mg, 20%) and the previously described compounds ent-18,12 $\alpha$ ,17-triacetoxybeyer-15-ene<sup>18</sup> (6) (70 mg, 35%), ent- $1B,16\alpha,17$ -triacetoxyatis-13-ene<sup>13</sup> (7) (30 mg, 15%) and ent-18,17-diacetoxy-16 $\alpha$ -hydroxyatis-13-ene<sup>13</sup> (8) (20 mg, 10%) and the starting product (5) (20 mg, 10%).

# DIMESYLATION AND REARRANGEMENT OF PRODUCT l.-

Three products were isolated, after CC, when dimesylation of product 1 (150 mg) was attempted with MsCl/pyridine  $(0.12 \text{ mL})$  under conditions of reflux  $(24 \text{ h})$ :  $(E)$ -ent-18-acetoxy-17-mesyloxyatis-13,16diene (10) (60 mg, 40%), (Z)-ent-1ß-acetoxy-17mesyloxyatis-13,16-diene (11) (37 mg, 25%) and ent-1ßacetoxy-16ß-hydroxy-17-mesyloxyatis-13-ene (12) (20 mg, 14%). Product 10: mp : 123-25°C;  $[\alpha]_D$ -16° (CHCl<sub>3</sub>, c 1); IR  $\mu_{\text{max}}$  (CHCl<sub>3</sub>): 2950, 1720 and 1165 cm<sup>-1</sup>; <sup>1</sup>H NMR: see Table I; <sup>13</sup>C NMR: see Table III; MS, m/z (%): 423 ([M+1]<sup>+</sup>, 1%), 269 (48), 365 (100). Anal. Calcd for C<sub>23</sub>O<sub>5</sub>H<sub>34</sub>S: C, 65.40; H, 8.06. Found: C, 65.44; H, 8.12. Product 11: mp : 127-29°C;  $\alpha$ <sub>1</sub>, -8° (CHCl<sub>3</sub>, c 1); IR  $\mu_{\text{max}}$  (CHCl<sub>3</sub>): 2945, 1720 and 1160 cm<sup>-1</sup>; <sup>1</sup>H NMR: see Table I; <sup>13</sup>C NMR: see Table III; MS, m/z (%): 423 ([M + 1]<sup>+</sup>, 2%), 269 (43), 365 (100). Anal.Calcd for  $C_{23}O_3H_{34}S$ : C, 65.40; H, 7.58. Found: C, 65.38; H, 8.02. Product 12: Colorless gum;  $\alpha|_{\rm p}$  + 15° (CHCl<sub>3</sub>, c 1); IR  $\mu_{\rm max}$  (CHCl<sub>3</sub>): 3520, 2945, 1720 and 1170 cm<sup>-1</sup>; <sup>1</sup>H NMR: see Table I; <sup>13</sup>C NMR: see Table III; MS, m/z (%): 441 ( $[M+1]^+$ , 2%), 423 (10), 379 (17), 321 (14), 285 (42), 363 (100).

# ACID TREATMENT OF PRODUCT 2.-

Product 2 (150 mg) was dissolved in Cl<sub>2</sub>CH<sub>2</sub> (20 mL), washed with a diluted solution of HCl (20 mL, 0,1N) during a few minutes, neutralized with NaHCO<sub>3</sub>, extracted with CH<sub>2</sub>Cl<sub>2</sub>, dried over anhidrous  $NaSO<sub>a</sub>$  and the solvent evaporated. After CC, the following products were isolated: ent-1ß-acetoxy-16ßhydroxy-17-mesyloxyatis-13-ene (13) (37 mg, 15%), ent-1ß-acetoxy-12a-hydroxy-17-mesyloxy- $16(13)$  +12,11(12) +13-diabeobeyer-15-ene (32) (22 mg, 15%) and starting product (2) (60 mg, 40%). Product 13: mp : 174-76°C;  $[\alpha]_D$  +6.4° (CHCl<sub>3</sub>, c 1); IR  $\mu_{max}$  (CHCl<sub>3</sub>): 3505, 2944, 1718 and 1174 cm<sup>-1</sup>; <sup>1</sup>H NMR: see Table I; <sup>13</sup>C NMR: see Table III; MS, m/z (%): 441 ( $[M+1]$ <sup>+</sup>, 1%), 285 (37), 363 (100). Anal. Calcd for C<sub>2</sub>O<sub>6</sub>H<sub>x</sub>S: C<sub>0</sub>62.73; H, 8.18. Found: C, 62.54; H, 8.14. Product 32: Colorless gum;  $\alpha$ <sub>ln</sub> -14° (CHCl<sub>3</sub>, c 1); IR  $\mu_{\text{max}}$  (CHCl<sub>3</sub>): 3510, 1740 and 1245 cm<sup>-1</sup>; <sup>1</sup>H NMR: see Table II; MS, m/z (%): 441  $([M+1]^+, 1\%)$ , 366 (38), 383 (100).

MESYLATION OF ent-1B-ACETOXY-12a,17-DIHYDROXYATIS-13-ENE (14).- 100 mg of 14, isolated as indicated in reference 22, was treated with 0.035 mL of MsCl. Work-up proceeded in the usual form, and after CC, product (13) (90 mg, 90%) was obtained.

ACETOLYSIS OF PRODUCT 13.-

Product 13 (80 mg) was dissolved in AcOK/AcOH (3 mL) and stirred under reflux for 24 h. Workup proceeded in **the** usual manner, and after CC, product 4 (68 mg, 85%) was isolated. OXIDATION OF PRODUCT l.-

100 mg of 1 was oxidized with pyridinium dichromate<sup>22</sup>, to give, after CC, ent-1ß-acetoxy-17hydroxy-15-en-12-one (15) (85 mg, 85%). mp : 61-63°C; [ $\alpha$ ]<sub>D</sub> -3.63 (CHCl<sub>3</sub>, c 1); IR  $\mu_{\text{max}}$  (CHCl<sub>3</sub>): 3450, 1750,and 1730 cm"; 'H NMR: see Table I; 13C NMR: see Table III; MS, m/z (%): 361 ([M+ l]', 1%), 241 (89), 229 (93), 301 (100). Anal. Calcd for  $C_2O_4H_3$ : C, 73.33; H, 17.78. Found: C, 73.60; H, 8.67. MESYLATION OF PRODUCT 15.-

90 mg of product 15 was mesylated with MsCl (0.03 mL). The solution was kept at room temperature for 24 h as described above. After CC, ent-1B-acetoxy-17-mesyloxybeyer-15-en-12-one (16)

(80 mg, 90%) was isolated. mp : 133-35°C; [ $\alpha$ ]<sub>D</sub> +40° (CHCl<sub>3</sub>, c 1); IR  $\mu_{max}$  (CHCl<sub>3</sub>): 3075, 1748 and 1732 cm<sup>-1</sup>; <sup>1</sup>H NMR: see Table I; <sup>13</sup>C NMR: see Table III; MS, m/z (%): 439([M+1]<sup>+</sup>, 1%), 283 (48), 379 (100). Anal. Calcd for  $C_{23}O_6H_3S$ : C, 63.01; H, 7.76. Found: C, 62.96; H, 7.92. ACETOLYSIS OF PRODUCT 16.-

Product 16 (70 mg) was treated with AcOK/AcOH for 24 h under reflux, after which only 16 (66 mg, 95%) was isolated.

# REDUCTION OF PRODUCT 15.-

NaBH<sub>4</sub> (900 mg) was added to a stirred solution of product 15 (450 mg) in i-PrOH (25 mL) at room temperature for 30 min. The reaction mixture was acidified with dil. HCI, extracted with CH,CI,, dried with NaSO<sub>4</sub> and concentrated in vacuo. Chromatography on a silica gel column yielded ent-18acetoxy-128,17-dihydroxybeyer-15-ene (17) (360 mg,80%) and product 1 (90 mg, 20%). Product 17: mp : 180°C;  $\lbrack \alpha \rbrack_p$  +68° (CHCl<sub>3</sub>, c 1); IR  $\mu_{\text{max}}$  (CHCl<sub>3</sub>): 3525, 2946, 1720 and 1170 cm<sup>-1</sup>; <sup>1</sup>H NMR: see Table I; <sup>13</sup>C NMR: see Table III; MS, m/z (%): 363 ([M+1]<sup>+</sup>, 2%), 285 (55). Anal. Calcd for  $C_2O_4H_4$ : C, 72.93; H, 9.39. Found: C, 72.74; H, 9.43.

# MESYLATION OF PRODUCT 17.-

Treatment of 17 (220 mg) with MsCl (0.15 mL) under the same conditions as described above gave ent-18-acetoxy-128,17-dimesyloxybeyer-15-ene (18) (198 mg, 90%). mp : 185-87°C;  $[\alpha]_D$  + 39° (CHCl<sub>3</sub>, c 1); IR  $\mu_{\text{max}}$  (CHCl<sub>3</sub>): 2945, 1720 and 1175 cm<sup>-1</sup>; <sup>1</sup>H NMR: see Table I; <sup>13</sup>C NMR: see Table III; MS, m/z  $(\%)$ : 519 ([M+1]<sup>+</sup>, 1%), 459 (26), 362 (40), 284 (60), 79 (100). Anal. Calcd for C<sub>24</sub>O<sub>8</sub>H<sub>38</sub>S<sub>2</sub>: C, 55.60; H, 7.34. Found: C, 55.72; H, 7.36.

ACETOLYSIS OF PRODUCT 18.-

Product 18 (200 mg) was treated with AcOK/AcOH as indicated above, yielding four products after CC: (E)-ent-1B,17-diacetoxy-14(13)~12-abeobeyer-13(17),15-diene (19) (80 mg, 40%), (Z)-ent-l&17 diacetoxy-14(13)-12-abeobeyer-13(17),15-diene (20) (40 mg, 20%), (E)-ent-1ß-acetoxy-17-mesyloxy-14(13)+12\_abeobeyer-13(17),15\_diene(21) (30 mg, 15%) and (Z)-ent-18acetoxy-17-mesyloxy-14(13)+12 abeobeyer-13(17),15-diene (22) (30 mg, 15%). Product 19: mp : 141-43°C; [ $\alpha$ ]<sub>n</sub> +32.6° (CHCl<sub>3</sub>, c 1); IR  $\mu_{\text{max}}$  (CHCl<sub>3</sub>): 2945, 1718, 1163 and 1165 cm<sup>-1</sup>; <sup>1</sup>H NMR: see Table I; <sup>13</sup>C NMR: see Table III; MS, m/z  $(\%)$ : 387 ( $[M+1]^+$ , 2%), 327 (78), 267 (18), 285 (100). Anal. Calcd for  $C_2O_4H_4$ : C, 74.61; H, 8.81. Found: C,74.42; H, 8.97. Product 20: mp : 137-39°C;  $[\alpha]_D$  +7.3° (CHCl<sub>3</sub>, c 1); IR  $\mu_{\text{max}}$  (CHCl<sub>3</sub>): 2947, 170, 1160 and 1163 cm<sup>-1</sup>; <sup>1</sup>H NMR: see Table I; <sup>13</sup>C NMR: see Table III; MS, m/z (%): 387 ( $[M+1]$ <sup>+</sup>, 2%), 327 (55), 267 (15), 285 (100). Anal. Calcd for C<sub>24</sub>O<sub>4</sub>H<sub>34</sub>: C, 74.61; H, 8.81. Found: C, 74.24; H, 8.78. Product 21: mp : 128-30°C;  $[\alpha]_D$  42.8° (CHCl<sub>3</sub>, c 1); IR  $\mu_{\text{max}}$  (CHCl<sub>3</sub>): 2945, 1720, 1160 and 1162 cm<sup>-1</sup>; <sup>1</sup>H NMR: see Table I; <sup>13</sup>C NMR: see Table IV; MS, m/z (%): 423 ([M+1]<sup>+</sup>, 8%), 327 (12), 267 (80), 363 (100). Anal. Calcd for C<sub>23</sub>O<sub>2</sub>H<sub>34</sub>S: C, 65.40; H, 8.06. Found: C, 64.98; H, 8.44. Product 22: mp : 130-32°C; [ $\alpha$ ]<sub>D</sub> +4.5° (CHCI<sub>3</sub>, c 1); IR  $\mu_{\text{max}}$  (CHCI<sub>3</sub>): 2945, 1719, 1162 and 1165 cm<sup>-1</sup>; <sup>1</sup>H NMR: see Table I; MS, m/z (%): 423  $([M+1]^*, 6\%)$ , 327 (8), 267 (82), 363 (100). Anal. Calcd for  $C_{23}O_{14}H_{24}S$ : C, 65.40; H, 8.06. Found: C, 65.04; H, 8.40.

# OXIDATION OF PRODUCT 14.-

Product **14 (500** mg), dissolved **in** MeOH/H,O, was added a flask containing 500 mg NaIO, dissolved in 3 mL water (in an ice bath with magnetic stirring). Then 200 mg of NaHCO<sub>3</sub> was also added. A white precipitate of the nonsoluble product appeared and, after CC, ent-1ß-acetoxy-17-noratis-13-en-16one (23) (475 mg, 95%) was obtained. mp : 113-15°C;  $[\alpha]_D$  +2° (CHCl<sub>1</sub>, c 1); IR  $\mu_{\text{max}}$  (CHCl<sub>1</sub>): 2947, 2870, 1728 and 1243 cm<sup>-1</sup>; <sup>1</sup>H NMR: see Table II; <sup>13</sup>C NMR: see Table IV; MS, m/z (%): 331 ([M+1]<sup>+</sup>,

100%), 271 (82), 229 (11), 273 (26). Anal. Calcd for  $C_{21}O_3H_{30}$ : C, 76.36; H, 9.09. Found: C, 76.15; H, 8.99. REDUCTION OF PRODUCT 23.-

Product 23 (200 mg) was dissolved in EtOH (8 mL) and 100 mg NaBH<sub>4</sub> was added. The mixture was stirred at room temperature for 2 h. After CC, ent-1B-acetoxy-16B-hydroxy-17-noratis-13-ene (24) (130 mg,  $65\%$ ) and ent-1ß-acetoxy-16 $\alpha$ -hydroxy-17-noratis-13-ene (25) (70 mg, 35%), were isolated. Product 24: mp : 135-36°C;  $[\alpha]_D$  + 2.5° (CHCl<sub>3</sub>, c 1); IR  $\mu_{max}$  (CHCl<sub>3</sub>): 3343, 2929, 2871, 2850 and 1245 cm<sup>-1</sup>; <sup>'</sup>H NMR: see Table II; <sup>13</sup>C NMR: see Table IV; MS, m/z (%): 333 ([M+1]<sup>+</sup>, 1%), 273 (59), 256 (16), 228 (20), 78  $(33)$ , 255 (100). Anal. Calcd for  $C_{21}O_3H_{32}$ : C, 75.90; H, 9.64. Found: C, 75.67; H, 9.50. Product 25: mp : 164-65°C;  $[\alpha]_D$  + 1.3° (CHCl<sub>3</sub> c 1); IR  $\mu_{max}$  (CHCl<sub>3</sub>): 3343, 2940, 2871, 2870, 1728 and 1246 cm<sup>-1</sup>; <sup>1</sup>H NMR: see Table II; <sup>13</sup>C NMR: see Table IV; MS, m/z (%): 333 ([M+1]<sup>+</sup>, 1%), 273 (50), 256 (20), 228 (22), 78 (35), 255 (100). Anal. Calcd for  $C_{21}O_{3}H_{32}$ : C, 75.90; H, 9.64. Found: C, 75.73; H, 9.54. MESYLATION AND REARRANGEMENT OF PRODUCT 24.-

Product 24 (100 mg) was treated with MsCl (0.02 mL) at room temperature in the usual manner to give, after CC, ent-18-acetoxy-12a-hydroxy-16(13)-12,11(12)-13-17-norbeyer-15-ene (26) (95 mg, 95%). mp : 129-30°C;  $[\alpha]_D$  + 2° (CHCl<sub>3</sub>, c 1); IR  $\mu_{max}$  (CHCl<sub>3</sub>): 3323, 2948, 2868, 1736 and 1240 cm<sup>-1</sup>; <sup>1</sup>H NMR: see Table II; <sup>13</sup>C NMR: see Table IV; MS, m/z (%): 333 ([M+1]<sup>+</sup>, 1%), 273 (34), 257 (41), 217 (36), 107 (86), 91 (100). Anal. Calcd for C<sub>21</sub>O<sub>3</sub>H<sub>22</sub>: C, 75.90; H, 9.64. Found: C, 75.65; H, 9.68. MESYLATION AND REARRANGEMENT OF PRODUCT 25.-

Treatment of 25 (100 mg) with MsCl (0.02 mL) under the same conditions as described for product 24, gave, after CC, ent-1 $\beta$ -acetoxy-14R-hydroxy-17-nortrachylobane (27) (85 mg, 85%). mp : 162-64°C; [ $\alpha$ ]<sub>D</sub> + 1.5° (CHCl<sub>3</sub>, c 1); IR  $\mu_{\text{max}}$  (CHCl<sub>3</sub>): 3484, 2932, 2871, 1717 and 1269 cm<sup>-1</sup>; <sup>1</sup>H NMR: see Table II; <sup>13</sup>C NMR: see Table IV; MS, m/z (%): 333 ([M+l]+, 2%), 273 (71), 256 (21), 228 (24), 255 (100). Anal. Calcd for  $C_{21}O_3H_{32}$ : C, 75.90; H, 9.64. Found: C, 75.98; H, 9.42. REDUCTION OF PRODUCT 23 WITH NaBD<sub>4</sub>.

Treatment of product 23 (200 mg) with  $NABD<sub>4</sub>$  under the same conditions as in treatment with NaBH<sub>4</sub> gave, after CC: ent-1B-acetoxy-16B-hydroxy-16a-deuterio-17-noratis-13-ene (28) (130 mg, 65%) and ent-1ß-acetoxy-16α-hydroxy-16ß-deuterio-17-noratis-13-ene (29) (70 mg, 35%). Product 28: mp : 144-45°C;  $[\alpha]_n$  + 2.3° (CHCl<sub>3</sub>, c 1); IR  $\mu_{\text{max}}$  (CHCl<sub>3</sub>): 3328, 2926, 2871, 1728 and 1246 cm <sup>-1</sup>; <sup>1</sup>H NMR: see Table II; <sup>13</sup>C NMR: see Table IV; MS, m/z (%): 334 ([M+1]<sup>+</sup>, 1%), 316 (12), 274 (70), 257 (20), 256 (100). Product 29: mp : 171-72°C;  $[\alpha]_D$  + 1.5° (CHCl<sub>3</sub>, c 1); IR  $\mu_{max}$  (CHCl<sub>3</sub>): 3484, 2945, 2853, 1718 and 1241 cm<sup>-1</sup>; <sup>1</sup>H NMR: see Table II; <sup>13</sup>C NMR: see Table IV; MS, m/z (%): 334 ([M+1]<sup>+</sup>, 1%), 316 (11), 275 (18), 257 (20) 256 (100).

## MESYLATION AND REARRANGEMENT OF PRODUCT 28.-

Mesylation of product 28 (100 mg) under the same conditions that described above for product 24 gave, after CC, ent-1ß-acetoxy-12 $\alpha$ -hydroxy-13-deuterio-16(13)-12,11(12)-13-diabeo-17-norbeyer-15-ene (30) (90 mg, 90%). mp : 145-47°C;  $[\alpha]_D$  +2.2° (CHCl<sub>3</sub>, c 1); IR  $\mu_{max}(CHCl_3)$ : 3403, 2944, 2870, 1736 and 1245 cm-'; 'H NMR: see Table II; 13C NMR: see Table IV; MS, m/z (%): 334 ([M+ l]', l%), 316 (45), 257 (22), 256 (96), 274 (100).

## MESYLATION AND REARRANGEMENT OF PRODUCT 29.-

Treatment of product 29 (100 mg) in the usual manner with MsCl gave, after CC, ent-1ß-acetoxy-14R-hydroxy -16-deuterio- 17-nortrachylobane (31) (80 mg, 80%). mp : 127-29°C;  $[\alpha]_D$  +9.8° (CHCl<sub>3</sub>, c 1); IR  $\mu_{\text{max}}$  (CHCl<sub>3</sub>): 3439, 2918, 2853, 1735 and 1236 cm<sup>-1</sup>; <sup>1</sup>H NMR: see Table II; <sup>13</sup>C NMR: see Table IV; MS, m/z (%): 334 ([M+ l]', 4%) 316 (20) 275 (20) 257 (15) 256 (60), 274 (100).

## DEHYDROXYIATION OF PRODUCI **l.-**

Product 1 (300 mg) was firstly treated with  $Py/Cl<sub>a</sub>C/Ph<sub>1</sub>P$  and afterward with tributyltin hydride/ $\alpha, \alpha'$ -azo-isobutyronitrile/toluene as described in references 23 and 24. After CC, ent-1ß-acetoxy- $12\alpha$ -hydroxybeyer-15-ene (33) (225 mg, 85%), was obtained. mp : 135-37°C;  $\alpha_{\text{ln}}$  +48.5° (CHCl<sub>3</sub>, c 1); IR  $\mu_{max}$  (CHCI<sub>3</sub>): 3448, 2947, 2868, 1735 and 1245 cm<sup>-1</sup>; <sup>1</sup>H NMR: see Table II; <sup>13</sup>C NMR: see Table IV; MS, m/z (%): 347 ( $[M+1]^+$ , 2%), 329 (46), 269 (98), 243 (30), 287 (100). Anal. Calcd for  $C_{22}O_3H_{34}$ : C, 76.30; H, 8.93. Found: C, 76.35; H, 9.89.

## MESYLATION OF PRODUCT 33.-

Product 33 (150 mg) was treated with MsCl in the usual manner to give the compounds previously described<sup>20</sup> (34) (90 mg, 60%) and (35) (30 mg, 20%) and *ent*-18-acetoxy-17-mesyloxyatis-13-ene (36) (22.5 mg, 15%). Product 36: Colorless gum;  $[\alpha]_D$  + 18.6° (CHCl<sub>3</sub>, c 1); IR  $\mu_{\text{max}}$  (CHCl<sub>3</sub>): 2931, 2842, 1732 and 1247 cm<sup>-1</sup>; <sup>1</sup>H NMR: see Table II; <sup>13</sup>C NMR: see Table IV; MS, m/z (%): 425 ([M+1]<sup>+</sup>, 2%), 366 (23), 329 (22), 270 (21), 269 (97), 365 (100).

# HYDROGENATION OF PRODUCT 2.-

Hydrogenation of 120 mg of 2, dissolved in 10 mL EtOH using  $Pd(5\%)/BaSO<sub>4</sub>$  as a catalyst, was carried out at room temperature and 5 atm. of  $H_2$  pressure for 2h. After CC, ent-1B-acetoxy-12a-hydroxy-17-mesyloxybeyerane (37) (120 mg, 100%), was isolated. Colorless gum;  $[\alpha]_D$  + 23° (CHCl<sub>3</sub>, c 1); IR  $\mu_{max}$ (CHCl<sub>3</sub>): 3528, 2946, 2871, 1731, 1249 and 1174 cm<sup>-1</sup>; <sup>1</sup>H NMR: see Table II; <sup>13</sup>C NMR: see Table IV; MS, m/z (%): 443 ( $[M+1]^+$ , 4%), 366 (24), 287 (33), 269 (25), 365 (100). Anal. Calcd for  $C_{23}O_6H_{38}S$ : C, 62.44; H, 8.60. Found: C, 62.88; H, 8.97.

ACETOLYSIS OF PRODUCT 37.-

Product 37 (90 mg) was treated with AcOK/AcOH as described above under General Acetolysis Procedure. After CC, ent-113,12a,17-triacetoxybeyerane (38) (72 mg, 80%). Product 38: mp : 136-38°C;  $[\alpha]_{D}$  +25.7° (CHCl<sub>3</sub>, c 1); IR  $\mu_{max}$  (CHCl<sub>3</sub>): 2948, 2871, 1735 and 1245 cm<sup>-1</sup>; <sup>1</sup>H NMR: see Table II; <sup>13</sup>C NMR: see Table IV; MS, m/z (%): 447 ([M+ l]', l%), 389 (88), 330 (23), 328 (15), 269 (40), 329 (100). Anal. Calcd for  $C_{16}O_6H_{38}$ : C, 69.96; H, 8.52. Found: C, 69.98; H, 8.48.

### ACKNOWLEDGEMENTS

We thank Karen Shashok for her assistance in the translation of the text. This work was supported by a grant from the Comisión Asesora de Investigación Científica y Técnica.

### REFERENCES

(1) Appleton, R.A.; M&lees, A.J.; McCormick, A.; McCrindle,R.; Murray, R.D. J. *Chem. Sot. C 1966, 2319.* 

- *(2)* Appleton, R.A.; **GUM,** P.A.; McCrindle, R.J. J. *Chem. Sot. C 1970, 1148.*
- *(3)* Coates, R.M.; Bertram, E.F. .I. Org. *Chem. 1971, 36, 3722.*
- *(4)* McAiees, A.J.; McCrindle, R. J. *Chem. Sot., Per&in* Trans. I 1975, 861.
- (5) McAlees, A.J.; McCrindle, R.; Murphy, S.T. J. *Gem. SOL, Perkin I* 1975, 1641.
- (6) Kapadi, A.H.; Dev, S. *Tetrahedron Left. 1965, 18, 1255.*
- *(7)* Hanson, JR. *Tetrahedron 1967, 23, 793.*
- *(8)* Yoshikoshi, A; Kitadani, M.; Kitahara, Y. *Tetrahedron 196?, 23, 1175.*
- 9. Buchanam, J.C.St.C.; Davis, B.R. *Chem. Common. 1967, 1142.*
- 10. Hanson, J.R. Tefrahedron 1970,26,2711.
- 11. Gunn, P.A.; McCrindle, R.; Roy, R.G. J. Chem. Soc. C 1971, 1018.
- 12. Murray, R.D.; Mills, R.W.; McAlees, A.J.; McCrindle, R. Tetrahedron 1974, 30,3399.
- 13. Ayer, WA.; Ball, J.H.; Rodriguez, B.; Valverde, S. Can. J. *Chem.* 1974, 52,2792.
- 14. Garcia-Granados, A.; Parra, A. *Anal. Quim.* 1983, 78C, 291.
- 15. Garcia-Granados, A.; Martinez, A.; Onorato, M.E. J. Org. Chem. 1987, 52, 606.
- 16. Laing, M.; Sommerville, P.; Hanouskova, D.; Pegel, K.H.; Piacenza, L.P.L.; Phillips, L.; Waight,
- E.S. J. C. S. *Chem. Comm. 1972,* 196.
- 17. Von Carstenn-Lichterfelde, C.; Valverde, S.; Rodriguez, B. *Aust. .I. Chem. 1974, 27,517.*
- 18. Escamilla, E.M.; Rodrfguez, B. *Phytochemistry 1980, 19,463.*

*19.* Jackmann, L.M.; Stlemhell, *S'ilpplication of NMR Spectroscopr in Organic Chemistry"* Pag. 281, Pergamon Press 2<sup>ª</sup> Ed., Oxford, 1969.

- 20. Cabrera, E.; García-Granados, A.; Hervás, M. J. Nat. Prod. 1989, 52, 669.
- 21. Garcia-Granados, A.; Parra, A.; Saenz de Buruaga, A.; Saenz de Buruaga, J.M. *An. Quim.* 1981, *77c,* 19.
- *22.* Corey, E.J.; Schidt, G. *Tetrahedron Letters 1979, 399.*
- *23.* Kuivila, H.G. *Synfhesk* 1970,499.
- 24. Downie, I.M.; Holmes, J.B.; Lee, J.B. Chem. Ind. 1966, 900.
- 25. Campbell, H.M.; Gunn, PA.; M&lees, A.J.; McCrindlc, R. *Can. J. Chem* 1975, 53,20.